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## Supramolecular Chemistry

Publication details, including instructions for authors and subscription information: <http://www.informaworld.com/smpp/title~content=t713649759>

# The Molecular Recognition of Amines with Calix[6]arene: Conclusive Xray and NMR Evidence for Endo and Exo Complex Formation between Calix[6]arene and Amines

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To cite this Article Lee, Young Ju , Park, Ki Deok , Yeo, Hyoung Min , Ko, Seung Whan , Ryu, Byung Ju and Nam, Kye Chun(2007) 'The Molecular Recognition of Amines with Calix[6]arene: Conclusive X-ray and NMR Evidence for Endo and Exo Complex Formation between Calix[6]arene and Amines', Supramolecular Chemistry, 19: 3, 167 — 173 To link to this Article: DOI: 10.1080/10610270600932818

URL: <http://dx.doi.org/10.1080/10610270600932818>

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# The Molecular Recognition of Amines with Calix[6]arene: Conclusive X-ray and NMR Evidence for Endo and Exo Complex Formation between Calix[6]arene and Amines

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(Received 23 May 2006; Accepted 9 June 2006)

Calix[6]arene formed an endo-complex with piperidine and its inclusion properties were investigated by NMR spectroscopy and X-ray crystallography. Especially, the DOSY spectrum showed that piperidine formed a unique complex with calix[6]arene.

Keywords: Calix[6]arene; Amine recognition; Exo complex; Endo complex; DOSY

#### INTRODUCTION

Ammonium ions play a fundamental role in a wide range of chemical and biological processes [1–4], and some are crucial in the medical field [5–7], which makes the recognition and sensing of amines a matter of great interest.

Since the first report of Gutsche's basic work on the interaction between calixarenes and amines [8], several papers have dealt with the thermodynamic and electrochemical aspects of these interactions [9– 11]. Some X-ray crystallographic studies have been reported about the structure and the mode of complexation. Thuery et al. [12] reported the X-ray structure of the doubly deprotonated form of calix[6]arene with two protonated triethylamines as counter-ions in a distorted 1,2,3-alternate conformation. Recently, Nachtigall et al. [13] studied the interactions of t-butylcalix[6]arene with aliphatic amines by  ${}^{1}$ H NMR and spectrophotometric titrations, and showed that calix[6]arene could be doubly deprotonated by aliphatic amines. Moreover, they reported that the extent of the second proton transfer was governed by the size of the substituent

The interaction between calixarene and amines proceeds by the transfer of a proton from the calixarene to the amine, followed by the formation of an *endo*-calixarene complex as evidenced by the  ${}^{1}H$ NMR spectroscopy. The apparent value of the chemical shift observed in such cases corresponds to the sum of the downfield shift induced by ion formation and the upfield shift caused by cation- $\pi$ interaction. The stability of the conformations of unsubstituted calixarenes is determined by intramolecular hydrogen bond formation [14]. Only two phenolic groups have ionizable protons and these have pKa values of 4.76 and 3.44, respectively [15].

Herein, we report on the binding properties of calix[6]arene and various amines based on NMR spectroscopy and X-ray structure analysis. Piperidine formed a strong endo-complex with calix[6]arene due to the strong basicity and a compact shape.

#### RESULTS AND DISCUSSION

## ${}^{1}$ H NMR and  $T_{1}$  Analysis

In order to investigate the interaction between calix[6]arene and amines,  $^{1}$ H NMR analysis was carried out. Figure 1 shows the  ${}^{1}$ H NMR spectra of

of the amine. The X-ray crystal structure of the dihexylammonium complex of the  $p$ -t-butylcalix[6] arene dianion confirmed its 1,2,3-alternate conformation and showed that one of the two ammonium moieties was encapsulated in the inner cavity of the calix[6]arene.

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ISSN 1061-0278 print/ISSN 1029-0478 online q 2007 Taylor & Francis DOI: 10.1080/10610270600932818



FIGURE 1  $^{-1}$ H NMR spectra of tripropylamine (a) and piperidine (b) in the presence of calix[6]arene in CDCl3. The numbers in the middle part indicate the number of molar equivalents of calixarene added.

tripropylamine and piperidine in the presence of calix[6]arene. The triplet at  $\delta$  2.37 ppm from methylene resonance of tripropylamine (Fig. 1a) was shifted downfield in the presence of calix[6]arene. On the other hand, the two broad singlets at  $\delta$  2.80 ppm and 1.52 ppm from piperidine (Fig. 1b) were shifted upfield considerably in the presence of calix[6]arene. Proton transfer from calix[6]arene to the amine forms an ammonium salt which makes the  $^1\mathrm{H}$  NMR resonance shift downfield as shown in Fig. 1a, suggesting that the tripropylamine salt forms an exo-complex with calix[6]arene. However, the large upfield shift of the proton resonance of piperidine in the presence of calix[6]arene indicates that the piperidine salt forms an endo-complex with calix[6]arene.

The chemical shifts of the various amines in the presence of calix[6]arene are summarized in Table I. A large upfield shift was observed in the case of piperidine  $(\Delta \delta - 0.42 \text{ ppm}$  and  $-0.60 \text{ ppm}$ , cyclohexylamine ( $\Delta \delta$  - 0.57 ppm), *n*-butylamine ( $\Delta \delta$  - 0.49 ppm,  $-0.27$  ppm and  $-0.18$  ppm), and DABCO ( $\Delta\delta$  $-0.21$  ppm), indicating that when these amines were used as the guest, they were embedded inside the cavity. This proves that the endo-type complexes are formed by  $CH-\pi$  interactions. A slight upfield shift was observed in the case of *t*-butylamine  $(\Delta \delta - 0.12)$ ppm), morpholine  $(\Delta \delta - 0.06$  ppm and  $-0.03$  ppm), and diethylamine ( $\Delta \delta$  – 0.07 ppm and – 0.15 ppm). In the case of diallylamine, and pyridine, the chemical shifts did not change at all. Downfield shifts were observed from triethylamine  $(\Delta \delta + 0.16 \text{ ppm}$  and 0.15 ppm) and tripropylamine  $(\Delta \delta + 0.10 \text{ ppm})$ , 0.06 ppm and 0.02 ppm). Obviously, it is necessary for the amines to have a certain basicity in order for them to withdraw protons from calix[6]arene. When the amine salts are formed, upfield or downfield shifts should be observed depending on whether exo- or endo-complexes are formed, respectively.

Figure 2 shows the relationship between amine basicity (pKa) and change in the chemical shift representing that amine basicity needs to be higher than pKa value of 10 for the formation of amine salts, except in the case of DABCO ( $pKa = 8.7$ ). Exocomplexes were formed from triethylamine and tripropylamine, whereas endo-complexes were formed from piperidine, cyclohexylamine, n-butylamine and DABCO, indicating that the shape (or size) of the amine determines the type of complex that is produced. The relatively compact shape of piperidine and cyclohexylamine due to their cyclic form made them the best fit for the calix[6]arene cavity. On the other hand, the bulkiness of triethylamine and tripropylamine prevented them from forming an endo-complex with calix[6]arene. Even though DABCO has a low basicity



TABLE I Chemical shift and relaxation time values for the protons in various amines in the presence of an equimolar amount of calix[6]arene in  $CDCl<sub>3</sub>$ 

\*Chemical shift ( $\delta$ ) and  $T_1$  were measured with the concentration of 10 mM at 27°C using 300 MHz NMR spectrometer. (a)  $\delta^{(a)}$  = Chemical shift of Amines in CDCl<sub>3</sub> (b)  $\delta^{(b)}$  = Chemical shift of Amines in the presence of Calix[6]arene in CDCl<sub>3</sub> (c)  $\Delta \delta = \delta^{(b)} - \delta^{(a)}$ .

 $(pKa = 8.7)$ , its compact shape allowed it to form an endo-complex with calix[6]arene.

Decreases in the <sup>1</sup>H spin lattice relaxation time  $(T_1)$ values were observed upon the complexation of calix[6]arene with the ammonium ions. This is consistent with a decrease in the mobility of the complexed calixarene in comparison with the free ligand. A significant decrease in the value of  $T_1$  was observed for piperidine (0.44 sec and 0.49 sec from 3.84 sec and 3.89 sec) when it was complexed with calix[6]arene, indicating that the endo-complex was formed. A decrease in the value of  $T_1$  observed for *n*butylamine was less than that of piperidine (0.81, 0.91 and 0.82 sec from 1.67, 1.84 and 2.45 sec). On the other hand, a relatively small decrease of  $T_1$  was observed for tripropylamine (1.23, 1.51 and 1.86 sec from 1.99, 2.42 and 2.61 sec), suggesting that the  ${}^{1}$ H

spin lattice relaxation was relatively slow due to the formation of the exo-complex of tripropylamine.

#### NOESY Analysis

The formation of the endo-complex can be confirmed by the 2D-NOESY spectrum of the calix[6]arenepiperidine complex. The correlation peaks between piperidine and the aromatic protons of calix[6]arene are shown in Fig. 3. The two peaks for piperidine (2.39 ppm and 0.92 ppm) were correlated with  $H_a$ ,  $H_b$ and  $H_c$  of calix[6]arene, clearly indicating that the endo-complex was formed with piperidine. However, in the case of triethylamine, no correlation peaks were observed between calix[6]arene and triethylamine in the NOESY anaysis.



FIGURE 2 Relationship between amine basicity and change in the chemical shift.



FIGURE 3 NOESY spectrum of the calix[6]arene-piperidine complex in CDCl<sub>3</sub>.

#### X-ray Structure Analysis

The crystal structure confirms that the calix[6]arene dianion exists as the 1,2,3-alternate conformation and the salt with the piperidinium ion. The molecular structure and perspective drawing of the complex are depicted in Fig. 4.

The calix[6]arene dianions form a complex with two piperidium cations for each calix[6]arene unit. One piperidium ion (N2) is placed in the cavity formed by three aromatic units of the 1,2,3-alternate conformation and binds the oxygens (O2, O6) of two other phenolic units by hydrogen bonds. On the other hand, the other piperidium ion (N1) sits outside the cavity and interacts only with one phenolate (O4) through a strong hydrogen bond. The detailed hydrogen bonding data for the X-ray structure are shown in Table II. A strong hydrogen bond was formed between the piperidium situated outside of the cavity and the phenolic hydroxyl group, so that a short distance of  $2.598 \text{ Å}$  was observed for  $N1-O4$ . On the other hand, two hydrogen bonds were formed between the endopiperidinium and calix[6]arene, corresponding to the  $N2$   $-$ O2 and  $N2$   $-$ O6 linkages with bond distances of 2.841 Å for 2.753 Å, respectively.



FIGURE 4 ORTEP projection of calix[6]arene dianion and piperidinium cations at the endo-calix (N2) and exo-calix (N1) <sup>1</sup><sup>1</sup><sup>1</sup></sub> positions. Hydrogen atoms involved in hydrogen bonds are only represented.

$D-HA$	$d(D-H)$	d(HA)	d(DA)	$Angle(D-HA)$
$N(2)$ -H(22N)O(6)	1.03(3)	1.86(3)	2.753(3)	143(2)
$N(2)$ -H(21N)O(2)	1.02(3)	1.91(3)	2.841(3)	150(3)
$N(1)$ -H(11N) $O(4)$	1.05(3)	1.56(3)	2.598(2)	171(2)
$O(6)$ -H(6O) $O(1)$	0.96(3)	1.54(3)	2.496(2)	175(3)
$O(5)$ -H(5O) $O(4)$	0.99(3)	1.59(3)	2.562(2)	169(3)
$O(3)$ -H(3O) $O(4)$	0.89(3)	1.71(3)	2.583(2)	168(3)
$O(2)$ -H(2O) $O(1)$	1.03(3)	1.49(3)	2.518(2)	173(3)

TABLE II Selected hydrogen bonds ( $\AA$ ) and angles ( $\degree$ ) in the crystal structure of dipiperidium calix[6]arene diphenolate

Symmetry transformations used to generate equivalent atoms.

#### **DOSY**

Diffusion-ordered NMR spectroscopy (DOSY), which utilizes stimulated spin echo (SPE) and its variants, separate the NMR signals of the different components in a mixture on the basis of their diffusion characteristics [16–19]. In this method, a series of spin-echo spectra are measured with different pulsed field gradient strengths, and the signal decays are analyzed in order to extract a set of diffusion coefficients. The 2D DOSY spectrum displays the conventional chemical shifts in one dimension and the diffusion coefficients in the other dimension. The diffusion dimension reveals the distribution of molecular sizes and allows different molecular species to be identified.

Therefore, we performed a DOSY experiment for a mixture of calix[6]arene, piperidine and tripropylamine in chloroform-d in order to examine the mobility of the host-guest complex. Figure 5a represents a DOSY spectrum of an equimolar mixture of piperidine and tripropylamine (each of 10 mM) and Fig. 5b shows a DOSY spectrum of the same mixture of amines with calix[6]arene (10 mM). When observing the mobility of a host and two guest molecules (Fig. 5b), it was found that the self diffusion coefficients of calix[6]arene and piperidine are almost the same, whereas tripropylamine shows significantly different diffusion coefficients suggesting supramolecular motion by a host-guest complex formed only from calix[6]arene and piperidine. A striking decrease in self diffusion coefficient of piperidine as the host-guest complex forms after addition of calix[6]arene can be seen by comparing two DOSY spectra. DOSY analysis is regarded as a powerful tool for molecular recognition in this case, because of showing its high selectivity for piperidine resulted from the shape of amine.

The chemical shift of piperidine (c) in the complex of calix[6]arene, piperidine and tripropylamine was upfield shifted as compared to the chemical shift of piperidine (b) in the 1:2 complex of calix[6]arene and piperidine as shown in Fig. 6. This is assumed to be due to the fact that the occupancy of the outer space by tripropylamine made it more likely for piperidine to exist in the cavity. The X-ray results showed that two piperidine molecules existed in the outer or inner cavity of the host. Meanwhile the NMR chemical shift of piperidine corresponded to the average value of the exo and endo forms, resulting from their interchange in the solution state.

#### EXPERIMENTAL

Calix[6]arene was prepared according to the known procedure.<sup>20</sup> The CDCl<sub>3</sub> used for the NMR experiments was purchased from Aldrich and used without further purification. For the relaxation measurement, NMR samples were prepared in 5 mm o.d. tubes, which were sealed under vacuum after degassing by several freeze-pump-thaw cycles. The <sup>1</sup>H NMR spectra were recorded at 300 MHz on a Varian Unity 300 spectrometer and the chemical shifts of the <sup>1</sup>H NMR spectra were referenced to TMS. The spin-lattice relaxation times  $(T_1)$  for protons were measured in  $CDCl<sub>3</sub>$  by means of the conventional inversion recovery method, using at least 16 different delays between  $\pi$  and  $\pi/2$  pulses, on a Varian Unity 300 spectrometer. Each  $T_1$  value was estimated with the aid of an exponential fitting program. The 2D experiments such as NOESY and DOSY, were performed on a Varian INOVA Unity-500 spectrometer equipped with a Performa II PFG system (up to  $60 \text{ G/cm}$  in the z-direction). The 2D-NOESY spectra were measured in the phasesensitive mode with mixing time and relaxation delay of 200 ms and 2.0 sec, respectively, at  $27^{\circ}$ C. The DOSY spectra were acquired with a single gradient ( $Z$ ) at  $25^{\circ}$ C using a 5 mm triple resonance indirect detection PFG probe, and measured by incrementing the amplitude of the field gradient pulses in 16 steps  $(0 \sim 46 \text{ Gcm}^{-1})$  using gcsteSL pulse sequence. The diffusion delay  $(\Delta)$  and the gradient pulse length  $(\delta)$ used were 50 ms and 2 ms, respectively.

Single crystals of the complex of bis-piperidinium calix[6]arene diphenolate were obtained by slow evaporation from chloroform. X-ray structure analysis was determined from data collection on a Bruker SMART CCD diffractometer equipped with graphite-monochromated Mo K $\alpha$  ( $\lambda = 0.71073$  A). The frame data were processed to give structure factors using the SAINT [21], and the structure was solved



FIGURE 5 DOSY spectra in CDCl<sub>3</sub>. (a) piperidine: tripropyl amine = 1:1, (b) piperidine: tripropylamine: calix[6]arene = 1:1:1.

by direct methods and refined by full-matrix leastsquares on  $F^2$  using SHELXTL software [22].

The positions of all hydrogen atoms including the hydrogen atoms on the ammonium(N1, N2) and phenolate (O2, O3, O5, O6) were found on the difference Fourier map and were refined with isotropic thermal displacement parameters.

Crystal data:  $C_{53}$  H<sub>59</sub> C<sub>13</sub> N<sub>2</sub> O<sub>6</sub>, M = 926.37 gmol<sup>-1</sup>,  $0.30 \times 0.25 \times 0.07$  mm, monoclinic,  $P2_1/c$ ,  $Z = 4$ , Absorption coefficient =  $0.245$  mm<sup>-1</sup>,  $a = 8.5458(4)$ Å,  $b = 28.0454(12)$  A,  $c = 19.9517(9)$  A,  $\alpha = 90^{\circ}$ ,  $\beta =$ 96.3470(10)°,  $\gamma = 90^{\circ}$ ,  $V = 4752.5(4)$   $\AA^3$ , 30410 reflections measured,  $R(int) = 0.0680$ ,  $R1 = 0.0480$ ,  $wR2 = 0.1013$ .



FIGURE 6 Comparison the chemical shift of piperidine in the  ${}^{1}$ H NMR spectra (a) piperidine (b) 1:2 calix[6]arene: piperidine (c) 1:1:1 mixture of calix[6]arene, piperidine and tripropylamine.

Crystallographic data for the structure reported in this article have been deposited with the Cambridge Crystallographic Data Center, CCDC No. 608666.

#### Acknowledgements

The authors are grateful to Professor S. S. Lee and Mr J. B. Suh at Gyeongsang National University and Professor M. S. Lah at Hanyang University for X-ray crystallography analysis. Korea Basic Science Institute, S. Korea is acknowledged for financial support (KBSI-2006-N25053).

#### References

- [1] Qiao, C.; Ling, K.; Shepard, E. M.; Dooley, D. M.; Sayre, L. M. J. Am. Chem. Soc. 2006, 128, 6206.
- [2] Kirchner, N.; Zedler, L.; Mayerhoefer, T. G.; Mohr, G. J. Chem. Comm. 2006, 1512.
- [3] Lu, H.; Grossman, J. E.; Lambert, J. B. J. Org. Chem. 2006, 71, 1769.
- [4] Colera, M.; Costero, A. M.; Gavina, P.; Gil, S. Tetrahedron: Asymmetry 2005, 16, 2673.
- [5] Xiao, J.; Guo, Z.; Guo, Y.; Chu, F.; Sun, P. Protein Eng. Des. Sel. 2006, 19, 47.
- [6] Achilias, D. S.; Sideridou, I. J. Macromol. Sci. Pure Appl. Chem. 2002, A39, 1435.
- [7] Ostah, N.; Lawson, G.; Zafar, S.; Harrington, G.; Hicks, G. J. Analyst 2000, 125, 111.
- [8] Bauer, L. J.; Gutsche, C. D. J. Am. Chem. Soc. 1985, 107, 6063.
- [9] De Namor, A. F. D.; Pardo, M. T.; Tanaka, D. A.; Velarde, F. J. S.; Garcia, J. D. J. Chem. Soc. Faraday Trans. 1993, 89, 2727.
- [10] De Namor, A. F. D.; Wang, J.; Orellana, J. G.; Velarde, F. J. S.; Tanaka, D. A. P. J. Incl. Phenom. Mol. Recogn. Chem. 1994, 19, 371.
- [11] Nachtigall, F. F.; Lazzarotto, M.; Nome, F. J. Braz. Chem. Soc. 2002, 13, 295.
- [12] Thuery, P.; Keller, N.; Lance, M.; Vigner, J. D.; Nierlich, M. J. Incl. Phenom. 1994, 20, 89.
- [13] Nachtigall, F. F.; Lazzarotto, M.; Castellano, E. E.; Nome, F. Supramol. Chem. 2004, 16, 453.
- [14] Atwood, J. L.; Clark, D. L.; Juneja, R. K.; Orr, G. W.; Robinson, K. D.; Vincent, R. L. J. Am. Chem. Soc. 1992, 115, 7558.
- [15] Gutsche, C. D.; Iqbal, M.; Alam, I. J. Am. Chem. Soc. 1987, 109, 4314.
- [16] Johnson, Jr., C. S. Prog. Nucl. Magn. Reson. Spectrosc. 1999, 34, 203.
- [17] Lin, M.; Shapiro, M. J.; Wareing, J. R. J. Am. Chem. Soc. 1997, 119, 5249.
- [18] Diaz, M. D.; Berger, S. Carbohydr. Res. 2000, 329, 1.
- [19] Stchedroff, M. J.; Kenwright, A. M.; Morris, G. A.; Nilsson, M.; Harris, R. K. Phys. Chem. Chem. Phys. 2004, 6, 3221.
- [20] Gutsche, C. D. In Monographs in Supramolecular Chemistry; Stoddart, J. F., Ed.; Royal Society of Chemistry: Cambridge, 1989, p 17.
- [21] Bruker, SMART and SAINT, Area Detector Control and Integration Software Ver.5.0; Bruker Analytical X-ray Instruments: Madison, Wisconsin, 1998.
- [22] Bruker, SHELXTL, Structure Determination Programs Ver. 5.16; Bruker Analytical X-ray Instruments: Madison, Wisconsin, 1998.